

# Effects of Pectin Molecular Weight Changes on the Structure, Dynamics, and Polysaccharide Interactions of Primary Cell Walls of *Arabidopsis thaliana*: Insights from Solid-State NMR

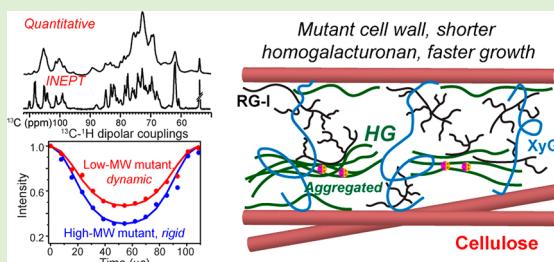
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 Supporting Information

**ABSTRACT:** Significant cellulose–pectin interactions in plant cell walls have been reported recently based on 2D <sup>13</sup>C solid-state NMR spectra of intact cell walls, but how these interactions affect cell growth has not been probed. Here, we characterize two *Arabidopsis thaliana* lines with altered expression of the *POLYGALACTURONASE INVOLVED IN EXPANSION1* (*PGX1*) gene, which encodes a polygalacturonase that cleaves homogalacturonan (HG). *PGX1*<sup>AT</sup> plants overexpress *PGX1*, have HG with lower molecular weight, and grow larger, whereas *pgx1-2* knockout plants have HG with higher molecular weight and grow smaller. Quantitative <sup>13</sup>C solid-state NMR spectra show that *PGX1*<sup>AT</sup> cell walls have lower galacturonic acid and xylose contents and higher HG methyl esterification than controls, whereas high molecular weight *pgx1-2* walls have similar galacturonic acid content and methyl esterification as controls. <sup>1</sup>H-transferred <sup>13</sup>C INEPT spectra indicate that the interfibrillar HG backbones are more aggregated whereas the RG-I side chains are more dispersed in *PGX1*<sup>AT</sup> cell walls than in *pgx1-2* walls. In contrast, the pectins that are close to cellulose become more mobile and have weaker cross peaks with cellulose in *PGX1*<sup>AT</sup> walls than in *pgx1-2* walls. Together, these results show that polygalacturonase-mediated plant growth is accompanied by increased esterification and decreased cross-linking of HG, increased aggregation of interfibrillar HG, and weaker HG–cellulose interactions. These structural and dynamical differences give molecular insights into how pectins influence wall dynamics during cell growth.



## INTRODUCTION

Plant cell walls are composed of interconnected networks of polysaccharides that include cellulose, hemicelluloses, and pectins. Cellulose contains multiple  $\beta$ -(1,4)-glucan chains that are hydrogen-bonded in parallel to form microfibrils.<sup>1,2</sup> Hemicelluloses differ between primary and secondary cell walls and between different plant taxa.<sup>3</sup> The main hemicellulose in the primary walls of eudicots is xyloglucan (XyG), whereas hemicelluloses in the primary walls of commelinid monocots consist largely of glucuronoarabinoxylan (GAX) and mixed-linkage glucans.<sup>3,4</sup> Hemicelluloses have been hypothesized to interact strongly with cellulose and serve as mechanical tethers or adhesives between cellulose microfibrils.<sup>5–7</sup>

While cellulose and hemicelluloses are neutral polymers, pectins are acidic polysaccharides with or without side chains.<sup>8,9</sup> Homogalacturonan (HG) is a linear homopolymer of  $\alpha$ -(1,4)-linked galacturonic acid (GalA), whose C6 carboxyl is often esterified upon HG delivery to the cell wall. These methyl esters can be subsequently removed by pectin methyl-esterases, enabling pairs of carboxyl groups on two HG chains to coordinate calcium ions and form interchain cross-links, stiffening the wall.<sup>10</sup> Thus, the degree of HG esterification impacts the biomechanical properties of cell walls. However,

the exact relationships between HG esterification, calcium-cross-linked network formation, wall extensibility, and cell growth are not well understood.<sup>10,11</sup> For example, although calcium cross-linking of de-esterified HG chains would be expected to rigidify the cell wall and constrain cell growth, de-esterification has been reported to correlate with wall weakening and increased cell expansion.<sup>10,12,13</sup> This suggests that de-esterified HG might be more susceptible to pectin-degrading enzymes such as polygalacturonases, which hydrolyze HG, or pectate lyases, which cleave HG via  $\beta$ -elimination.<sup>8</sup> The second major pectin in primary cell walls is rhamnogalacturonan I (RG-I), which contains a backbone with alternating rhamnose (Rha) and GalA units and neutral side chains that consist of arabinan, galactan, and arabinogalactan.<sup>14,15</sup> RG-I side chains have been hypothesized to mediate pectin interactions with cellulose.<sup>16</sup> The third type of pectin is rhamnogalacturonan II, which contains an HG backbone decorated with multiple complex side chains and can be cross-linked via borate diesters.<sup>14</sup>

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For decades, the three-dimensional structures of plant cell walls have been difficult to determine due to the insoluble and heterogeneous nature of cell walls. As a result, it is not well understood how cellulose and matrix polysaccharides interact noncovalently to form a strong but extensible network for plant cell stability and growth. Earlier models of eudicot primary walls hypothesized that XyG coats the surface of cellulose microfibrils and tethers them into a load-bearing network, which is embedded in a second phase formed by well-hydrated pectins.<sup>6,7,17,18</sup> However, recent biochemical and biomechanical measurements and solid-state NMR data suggest an alternative model in which XyG is entrapped at a small number of sites between cellulose microfibrils to regulate wall loosening.<sup>19–21</sup> This biomechanical hotspot model is manifested in terms of a limited number of XyG–cellulose cross peaks in 2D and 3D <sup>13</sup>C correlation NMR spectra, binding of the wall-loosening protein,  $\alpha$ -expansin, to the XyG-enriched region of the cellulose microfibril,<sup>22,23</sup> and changes in cellulose microfibril synthesis and organization in the XyG-deficient *xxt1 xxt2* mutant.<sup>24</sup>

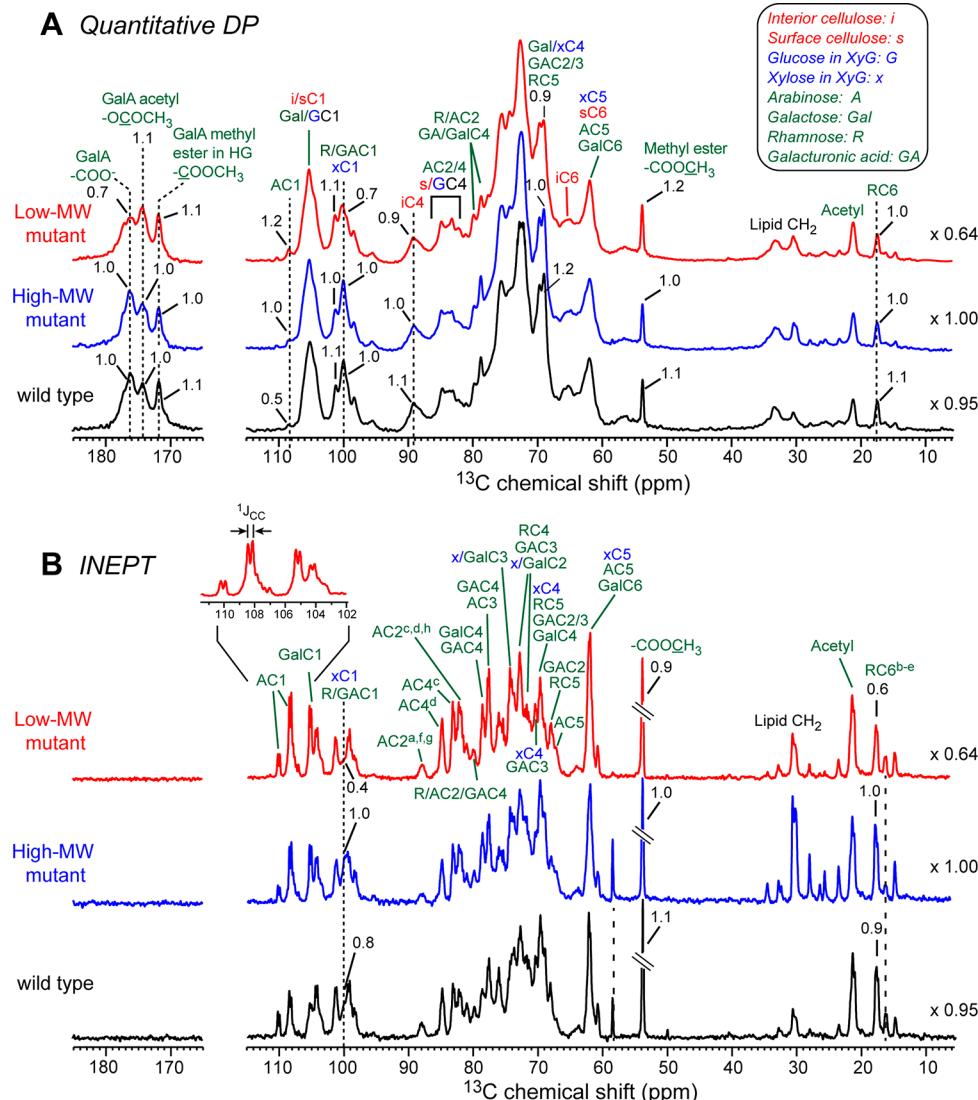
We have recently employed <sup>13</sup>C-enriched whole plants and multidimensional <sup>13</sup>C correlation solid-state NMR spectroscopy to investigate the structure and dynamics of polysaccharides in intact primary cell walls. Eudicot and commelinid monocot plants, including *Arabidopsis thaliana*, *Brachypodium distachyon*, and *Zea mays*, have been studied.<sup>25,26</sup> In eudicot cell walls, intermolecular cross peaks in 2D and 3D <sup>13</sup>C correlation spectra indicated that cellulose microfibrils are within a nanometer of both pectins and XyG; thus, the three major polymers form a single interconnected network.<sup>22,27</sup> In particular, a significant number of cellulose-HG and cellulose-RG I cross peaks were observed, indicating that cellulose and pectins interact on the subnanometer scale.<sup>28,29</sup> <sup>13</sup>C spin-lattice relaxation times ( $T_1$ ) and <sup>1</sup>H rotating-frame spin-lattice relaxation times ( $T_{1\rho}$ ) show a bimodal distribution of relaxation times for pectins, indicating that pectin backbones have a mobile domain and a rigid domain, which may correspond to segments that reside between cellulose microfibrils and segments that interact with cellulose, respectively.<sup>28</sup> Removal of calcium-cross-linked HG decreased the amount of bound water and slowed water <sup>1</sup>H spin diffusion to both pectins and cellulose,<sup>30</sup> which are also consistent with the model that a sizable fraction of the cellulose microfibril surface is coated by pectins.

These SSNMR data suggest a cell-wall model that highlights limited but mechanically important XyG–cellulose interactions and extensive pectin–cellulose spatial contacts.<sup>26,31</sup> The significant pectin–cellulose interactions were unexpected for two reasons. First, they appear at odds with *in vitro* binding assays that show that pectins have much lower binding capacity than XyG for cellulose.<sup>32–35</sup> For example, arabinan-rich beet pectins exhibit a binding capacity of  $\sim 10$   $\mu\text{g}/\text{mg}$  cellulose, whereas the XyG binds cellulose at  $\sim 120$   $\mu\text{g}/\text{mg}$  cellulose.<sup>36</sup> Only XyG–cellulose junctions bind expansins and respond to endoglucanase digestions, whereas pectin digestion does not induce significant creep extension in isolated wall samples.<sup>21</sup> However, the apparent discrepancies between binding assays and SSNMR-detected pectin–cellulose cross peaks may be reconciled if pectins were entrapped within each microfibril or between microfibrils during synthesis, transport, and assembly of the cell wall, a process that is difficult to replicate *in vitro*. Indeed, recent studies of cellulose synthesis and hydrogel formation in the presence of pectin solutions found that 20–40% of pectins interact intimately with cellulose during

assembly, as manifested by increased microfibril diameters in small-angle X-ray scattering data, denser networks of cellulose microfibrils in scanning electron micrographs, and altered viscoelastic and deformation properties of pectin–cellulose hydrogels compared to pure cellulose.<sup>37,38</sup>

A second reason for the skepticism about extensive pectin–cellulose interactions in cell walls is that it has not yet been shown whether these interactions are important for wall mechanics,<sup>21,23</sup> i.e., if perturbing the pectin–cellulose interactions can result in cell walls with altered mechanical and growth properties. Mutagenesis has been a valuable tool for understanding the structure–function relationships of cellulose and XyG in plant cell walls. For example, mutations in *CELLULOSE SYNTHASE* genes, *cesa1<sup>ageus</sup>* and *cesa3<sup>ixr1-2</sup>*, in *Arabidopsis* were shown by SSNMR and X-ray diffraction to disrupt cellulose organization, causing microfibrils with substantially lower crystallinity and an additional conformation of glucan chains.<sup>39</sup> *xxt1xxt2* mutants that lack XyG<sup>40</sup> were found to exhibit much shorter <sup>13</sup>C and <sup>1</sup>H relaxation times for cellulose and pectins compared to those of wild-type controls, indicating that XyG depletion increases the mobility of the remaining wall polysaccharides.<sup>22</sup> Another cellulose-deficient mutant, *cesa7<sup>ixr3-7</sup>*, was used to probe xylan–cellulose interactions in secondary cell walls. Whereas xylan flattens into a 2-fold helical screw conformation to bind to the surface of cellulose in wild-type secondary cell walls, xylan in the *cesa7<sup>ixr3-7</sup>* mutant lacks this 2-fold conformation.<sup>41,42</sup> Similar to these cellulose and XyG mutants, plants with altered pectin biosynthesis and processing should also be useful for elucidating how pectin–cellulose interactions impact wall structure and loosening.<sup>8,43</sup> Putative pectin biosynthetic mutants that are defective in extractable pectin content and esterification are known,<sup>44–48</sup> but several of these mutants also have altered xylan content. Recently, mutants overexpressing or lacking polygalacturonase genes have been shown to have altered HG molecular weights,<sup>43,49</sup> thus resulting in the possibility of characterizing cell walls with different pectin molecular weight profiles but potentially normal pectin biosynthesis.

In this study, we use SSNMR spectroscopy to examine the structures and mobilities of polysaccharides in two *Arabidopsis* mutant cell walls with altered pectin molecular weights. The *PGX1* gene encodes a polygalacturonase that hydrolyzes the HG backbone. A *PGX1<sup>AT</sup>* activation-tagged mutant over-expresses *PGX1* and has higher total polygalacturonase activity, resulting in an average HG molecular weight of  $\sim 66$  kDa,<sup>43</sup> which is lower than the wild-type HG molecular weight of  $\sim 150$  kDa. This *PGX1<sup>AT</sup>* mutant shows taller etiolated seedlings and larger adult plants than wild-type *Arabidopsis*. In comparison, a *pgx1-2* knockout mutant ablates *PGX1* expression, resulting in a higher HG molecular weight of  $\sim 200$  kDa. The knockout mutant produces shorter etiolated seedlings and smaller adult plants.<sup>43</sup> Using <sup>13</sup>C solid-state NMR, we show that the cell walls of these two mutants have altered pectin dynamics and pectin–cellulose interactions from each other and from the control cell wall. For the low-MW *PGX1<sup>AT</sup>* mutant, the dynamic pectin domain exhibits reduced HG and RG-I backbone mobilities but increased RG-I side chain mobilities, whereas the rigid pectin domain exhibits increased backbone mobility and weaker pectin–cellulose cross peaks in the low-MW mutant cell wall. These results suggest that pectins regulate the growth properties of plant cell walls by interacting with cellulose and/or by establishing an interfibrillar matrix



**Figure 1.**  $^{13}\text{C}$  MAS spectra of the low-MW (red) mutant, high-MW mutant (blue), and wild-type (black) *Arabidopsis* cell walls. Polysaccharide abbreviations for  $^{13}\text{C}$  resonance assignment are shown in the upper-right corner. (A) Quantitative  $^{13}\text{C}$  DP spectra plotted after multiplying the low-MW spectral intensities by 0.64-fold and the wild-type spectral intensities by 0.95-fold to account for sample amount differences. Intensity values for selected peaks indicate relative intensities among the three cell walls after taking into account sample amount differences. (B)  $^{13}\text{C}$  INEPT spectra showing only the signals of highly mobile polysaccharides.  $^{13}\text{C}$ – $^{13}\text{C}$   $J$  couplings can be resolved for some peaks, as shown, for example, for the Ara and Gal C1 peaks of the low-MW sample.

network, thus supporting the importance of pectin structure and pectin–cellulose interactions for controlling wall dynamics during cellular growth.

## MATERIAL AND METHODS

**Uniform  $^{13}\text{C}$  Labeling and Growth of Wild-Type and Mutant Cell Walls.**  $^{13}\text{C}$ -enriched *Arabidopsis thaliana* Columbia ecotype (Col-0), *PGX1*-overexpressed low-MW mutant (*PGX1<sup>AT</sup>*) and *PGX1*-depleted high-MW mutant (*pgx1-2*) primary cell walls were prepared as previously described.<sup>30,43</sup> The seedlings were grown in 2.2 g/L of Murashige and Skoog (MS) liquid culture containing 0.5%  $^{13}\text{C}$ -labeled glucose as the sole carbon source in the dark with shaking for 14 days before they were harvested. The  $^{13}\text{C}$  labeling level is estimated to be close to 100% based on cross peak intensities in 2D  $^{13}\text{C}$ – $^{13}\text{C}$  correlation spectra. The seedlings were ground into fine powder in liquid nitrogen, and the samples were washed with 1.5% (w/v) SDS for 3 h to solubilize cell membranes and proteins and inactivate endogenous wall-degrading enzymes. The material was thoroughly washed with water, incubated with  $\alpha$ -amylase (5,000 units per 30 mL)

from porcine pancreas (Sigma-Aldrich) in sodium MES buffer (pH 6.8) to remove starch and then digested with Pronase (200 units, 5 mg/20 mL, Sigma-Aldrich) in sodium MES buffer (pH 7.5) to digest proteins at 40 °C overnight. Incubation in 1.5% (w/v) SDS with shaking at room temperature for 1 h inactivated the Pronase. A total of 0.02% (w/v)  $\text{Na}_3\text{N}$  was used in all solutions to inhibit microbial growth. The samples were centrifuged to remove bulk water.<sup>30</sup>

**Solid-State NMR (SSNMR) Spectroscopy.** All SSNMR spectra were measured on an 800 MHz (18.8 T) and a 600 MHz (14.1 T) Bruker Advance II HD spectrometer using 3.2 mm magic-angle-spinning (MAS)  $^1\text{H}$ / $^{13}\text{C}$ / $^{15}\text{N}$  probes. Typical radiofrequency field strengths were 40–62.5 kHz for  $^{13}\text{C}$  and 50–80 kHz for  $^1\text{H}$ .  $^1\text{H}$  two-pulse phase modulation (TPPM) decoupling was applied during acquisition. All  $^{13}\text{C}$  chemical shifts were externally referenced to the adamantine  $\text{CH}_2$  peak at 38.48 ppm on the tetramethylsilane (TMS) scale.

1D  $^{13}\text{C}$  MAS spectra were measured at 296 K under 10 kHz MAS on the 800 MHz spectrometer using  $^1\text{H}$ – $^{13}\text{C}$  cross-polarization (CP) or  $^{13}\text{C}$  direct polarization (DP) to generate the initial magnetization. Quantitative  $^{13}\text{C}$  spectra were measured using  $^{13}\text{C}$  DP and a recycle

**Table 1.**  $^{13}\text{C}$  Chemical Shifts of Matrix Polysaccharides in the Low-MW Mutant (L), High-MW Mutant (H), and Wild-Type (WT) Cell Walls<sup>a</sup>

sugar	linkage	cell walls	C1 (ppm)	C2 (ppm)	C3 (ppm)	C4 (ppm)	C5 (ppm)	C6 (ppm)
Ara <sup>a</sup>	2,5-Ara	L, H, I	107.1	87.7				
Ara <sup>b</sup>	2,5-Ara	L, H, I	107.0	85.8				
Ara <sup>c</sup>	5-Ara	L, H, I, 2W, WT (all)	108.2	81.8	77.7	83.1	67.6	
Ara <sup>d</sup>	t-Ara	all	107.7	82.3	77.4	84.5	62.0	
Ara <sup>e</sup>		L, H, I, 2W	108.2	80.1				
Ara <sup>f</sup>	2,5-Ara	L, H, 2W, WT	108.5	88.0				
Ara <sup>g</sup>	2,5-Ara	L, H, WT	108.6	87.4				
Ara <sup>h</sup>	5-Ara	all	109.9	82.0				
Xyl <sup>a</sup>	t-Xyl of XyG	all	99.6	72.2	74.3	70.2	62.3	
Xyl <sup>b</sup>		H	99.0	72.5		69.3	62.4	
Gal <sup>a</sup>	4-Gal	all	105.1	72.8	74.0	78.4	75.2	61.5
Gal <sup>b</sup>	t-Gal	all	105.2	72.0	73.4	69.5	75.6	61.7
Gal <sup>c</sup>		L, H, I, 2W	104.1	72.6				
Gal <sup>d</sup>		all	103.4	73.8				
Gal <sup>e</sup>		all	104.1	71.4				
Gal <sup>g</sup>		L, H, WT	103.9	70.7				
GalA <sup>a</sup>	4-GalA	L, H, I, WT	99.8	69.0	69.7	78.5	72.1/71.2	
GalA <sup>b</sup>		all	101.0	68.6	71.3			
GalA <sup>c</sup>		all	100.1	69.5				
GalA <sup>g</sup>		L, H, 2W, WT	98.8	67.8	72.4	77.8	71.2	171.5
GalA <sup>i</sup>		L, H, 2W, WT	98.2	68.7	69.7	78.5	72.1	
Rha <sup>b</sup>		all				69.7	17.3	
Rha <sup>c</sup>		all				67.9	16.7	
Rha <sup>d</sup>	2-Rha	L, H, 2W, WT		79.8	71.0	72.6	68.1	16.0
Rha <sup>e</sup>		L, H, WT					68.4	17.4

<sup>a</sup>These chemical shifts are compared to previously reported polysaccharide chemical shifts found in 2 week old *Arabidopsis* seedling cell walls (2W) and inflorescence cell walls (I). Italicized chemical shifts are not resolved. Superscripts refer to subtypes of monosaccharides observed in this study. Some subtypes have been previously observed in the two week old and inflorescence cell walls but are absent in the high- and low-MW mutants. Very weak peaks such as Xyl-b in the low-MW cell wall are not included.

delay of 30 s.  $^{13}\text{C}$  CP spectra were measured using a contact time of 1 ms and a recycle delay of 2 s. Refocused INEPT<sup>50</sup> spectra were measured using a total polarization transfer time of 6 ms, which consists of two delays of 1.8 ms followed by two delays of 1.2 ms. These values correspond to  $1/(4J_{\text{CH}})$  and  $1/(6J_{\text{CH}})$ , respectively, assuming a standard  $^1\text{J}_{\text{CH}}$  value of 140 Hz.<sup>51</sup> A recycle delay of 3.5 s was used for the INEPT experiments.

2D  $^{13}\text{C}$ – $^{13}\text{C}$  J-INADEQUATE spectra<sup>52,53</sup> correlating the double- and single-quantum  $^{13}\text{C}$  chemical shifts were measured to assign the resonances of matrix polysaccharides. The experiments were carried out on the 800 MHz spectrometer at 296 K under 12 kHz MAS. To preferentially detect dynamic components of the cell wall, the initial  $^{13}\text{C}$  magnetization was generated using a  $^{13}\text{C}$  90° pulse and the recycle delay was 2 s. The spectral widths were 333 ppm (67 kHz) and 83 ppm (17 kHz) for the direct and indirect dimensions, respectively. A total of 320  $t_1$  increments were collected, and the number of scans per  $t_1$  increment was 128 for the low-MW mutant and wild-type cell walls and 256 for the high-MW mutant.

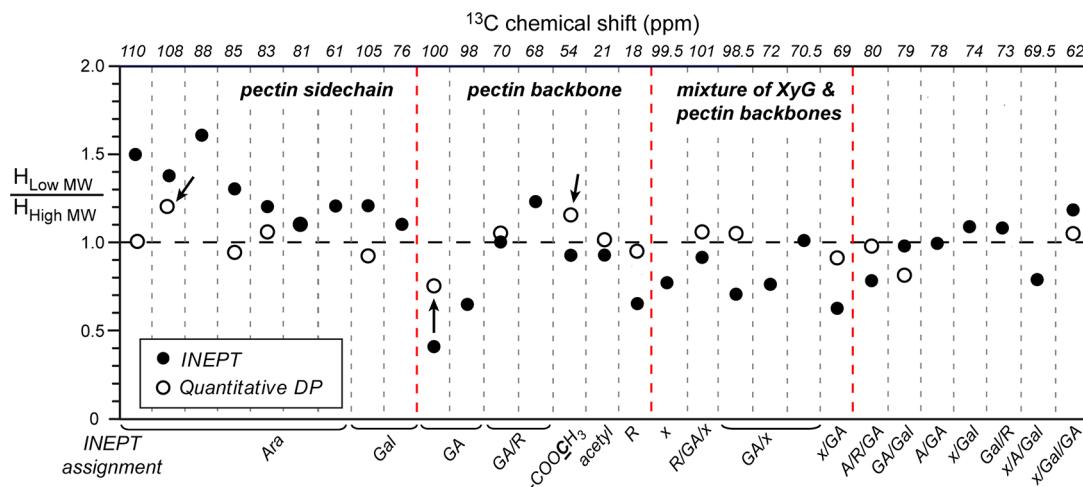
2D  $^1\text{H}$ -driven  $^{13}\text{C}$  spin diffusion (PDSD) spectra were measured on the 600 MHz spectrometer under 10 kHz MAS using a CP contact time of 300  $\mu\text{s}$ , a spin diffusion mixing time of 1.5 s, and a recycle delay of 1.3 s. The spectra were measured at 253 K to immobilize the pectins. The spectral widths were 331 ppm (50 kHz) and 110 ppm (17 kHz) for the direct and indirect dimensions; the  $t_1$  increments were 122 and 132 for the low- and high-MW mutants, respectively, and the corresponding numbers of scans were 192 and 352, respectively.

Dipolar-doubled  $^{13}\text{C}$ – $^1\text{H}$  dipolar chemical-shift correlation (DIPSHIFT) experiments were carried out on the 800 MHz spectrometer

at 296 K under 9.2 kHz MAS<sup>54,55</sup> using a recycle delay of 2.7 s.  $^1\text{H}$  homonuclear decoupling was achieved using the FSLG<sup>56</sup> pulse sequence with a  $^1\text{H}$  transverse field strength of 80 kHz. The FSLG scaling factor of 0.577 was verified using the model peptide formyl-Met-Leu-Phe-OH.<sup>57,58</sup> A 3:1 combination of dipolar couplings  $\delta$  and  $3\delta$  was used to simulate the DIPSHIFT curves of  $\text{CH}_3$  groups.<sup>55</sup> The ratios between the experimentally measured couplings and the effective rigid-limit C–H dipolar coupling of 26.2 kHz gave the order parameters  $S_{\text{CH}}$ ,  $S_{\text{CH}_2}$ , and  $S_{\text{CH}_3}$ .

## RESULTS

**Polysaccharide Compositions of Mutant Cell Walls from Quantitative  $^{13}\text{C}$  Spectra.** We first compared the polysaccharide compositions of wild-type and mutant cell walls using quantitative  $^{13}\text{C}$  spectra (Figure 1A). These spectra were measured using a recycle delay of 30 s, which is more than 5-times longer than the  $^{13}\text{C}$   $T_1$  relaxation times of 0.3–5 s for these uniformly  $^{13}\text{C}$ -labeled *Arabidopsis* cell walls.<sup>27,28</sup> These  $^{13}\text{C}$   $T_1$  relaxation times are significantly shorter than the  $T_1$ s of unlabeled cell walls<sup>59</sup> because  $^{13}\text{C}$  spin diffusion efficiently transfers magnetization from relaxation hotspots to all nearby  $^{13}\text{C}$  sites. The different cell walls show similar line widths in the quantitative  $^{13}\text{C}$  spectra, thus allowing us to use peak heights to report polysaccharide concentration differences among the three cell walls. To account for sample amount differences, we normalized the peak intensities by the integrated spectral



**Figure 2.** Quantitative and INEPT  $^{13}\text{C}$  intensity ratios between the low- and high-MW cell walls. The peak heights of the low-MW spectrum are scaled by 0.64 to account for sample amount differences with the high-MW spectrum. Most quantitative intensity ratios are close to 1, except for sites indicated by arrows, whereas INEPT intensity ratios deviate more significantly from 1, indicating significant differences in polysaccharide mobilities between the two cell walls.

intensities, excluding the 25–30 ppm lipid region. Relative to the high-MW spectrum, the low-MW mutant and wild-type spectra were scaled by a factor of 0.64 and 0.95, respectively.

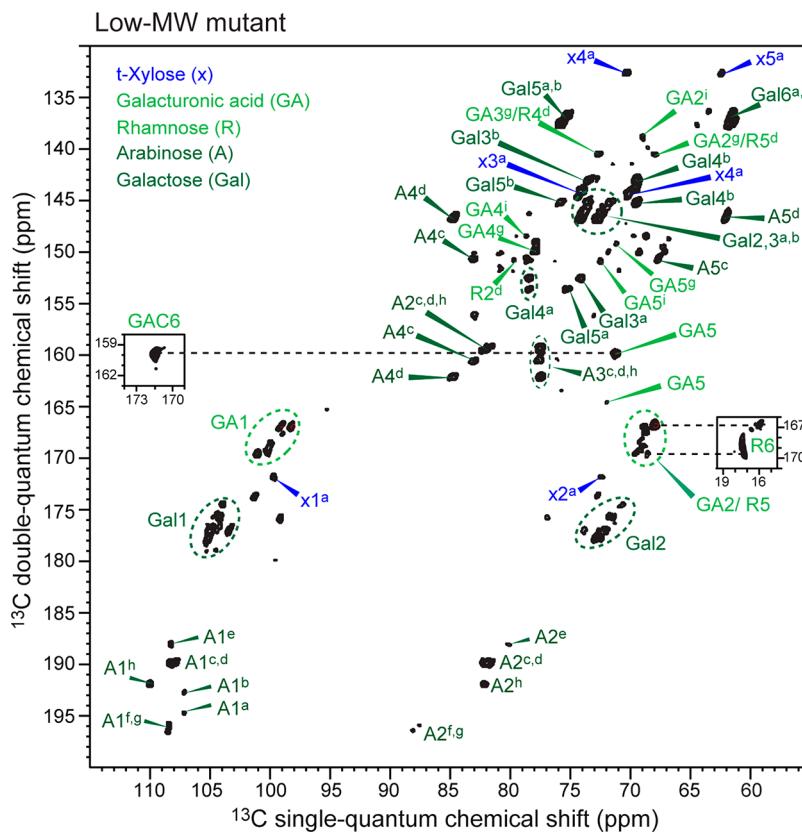
Careful inspection of the HG and RG-I peak intensities in the quantitative spectra gave detailed information about changes in methyl esterification and side chain decoration of the pectins among the three cell walls. First, the two mutants show much higher intensities of the 108 ppm arabinose (Ara) C1 signal than those of the wild-type cell wall, indicating that both mutants have higher concentrations of Ara side chains. Second, the wild-type and high-MW cell walls have the same intensities for the 100 ppm GalA and Rha C1 peak, the 174 ppm acetyl carbonyl peak, and the 176 ppm of the  $\text{COO}^-$  peak, whereas the low-MW cell wall exhibits different intensities. Specifically, the low-MW mutant has the same or higher intensities for the 101 ppm GalA and Rha C1 peak, the 174 ppm acetyl peak, and the 172 ppm methyl ester carbonyl peak, but lower intensities for the 100 ppm peak and the 176 ppm of  $\text{COO}^-$  peak. We can distinguish the assignment of the 100 and 101 ppm peaks based on their correlated intensity changes with the carbonyl peaks. Because the 176 ppm peak results from unesterified carboxyl groups, its intensity reduction along with the 100 ppm peak suggests that the 100 ppm peak can be assigned to C1 of unesterified and negatively charged GalA. Moreover, although GalA is present in both HG and RG-I, the 100 ppm peak can be specifically assigned to HG for the following reason. The Rha concentration has been previously found to be similar between wild-type and mutant cell walls of 6 day old dark-grown seedlings.<sup>43</sup> Consistently, our current 14 days old seedling samples show similar Rha C6 peak intensities for all three cell walls, indicating that the RG-I concentration is similar among the three cell walls. Therefore, the lower intensity of the 100 ppm peak in the low-MW cell wall can be attributed mainly to a lower concentration of the negatively charged HG compared to the high-MW and wild-type cell walls. Further supporting this conclusion, the 54 ppm methyl ester peak is higher in the low-MW spectrum than in the other two spectra, indicating a higher HG esterification level. Finally, the low-MW mutant shows a higher 21 ppm peak of  $\text{OCOCH}_3$  and 174 ppm peak of  $\text{OCOCH}_3$  than those of the other two cell walls, indicating a higher acetylation level. Taken together,

these observations indicate that the low-MW mutant has increased HG methyl esterification and acetylation and correspondingly lower amounts of negatively charged  $\text{COO}^-$  groups.

#### Dynamics of Matrix Polysaccharides from INEPT $^{13}\text{C}$ Spectra.

To selectively detect highly mobile polysaccharides, we measured  $^{13}\text{C}$  insensitive nuclei enhanced by polarization transfer (INEPT) spectra (Figure 1B). During  $J$  coupling-based  $^1\text{H}$ – $^{13}\text{C}$  polarization transfer, no  $^1\text{H}$ – $^1\text{H}$  homonuclear decoupling was applied; thus, the signals of rigid polysaccharides were suppressed by  $^1\text{H}$ – $^1\text{H}$  dipolar couplings.<sup>50</sup> The INEPT spectra show much higher resolution than the quantitative spectra:  $^{13}\text{C}$ – $^{13}\text{C}$   $J$  splittings can be resolved for some of the matrix polysaccharide signals, and the cellulose signals at 105, 89, 70–78, and 60–66 ppm are largely suppressed. The removal of the interior cellulose C4 peak revealed an 88 ppm Ara C2 peak. 2D  $^{13}\text{C}$  INADEQUATE spectra (see below) indicate that this 88 ppm peak can be assigned to Ara types a, f, and g (Table 1). Indeed, multiple Ara C2 and C4 peaks between 80 and 90 ppm are resolved and can be assigned to various subtypes based on the 2D  $J$ -INADEQUATE spectra. In addition, several GalA C1 peaks are resolved at 101 ppm (type b), 100 ppm (types a and e), 99 ppm (type g), and 98 ppm (type i). On the basis of the intensity ratios between the INEPT spectra and the quantitative spectra, and the expected 4-fold sensitivity enhancement by the INEPT polarization transfer, we estimate that these highly mobile polysaccharides account for less than 10% of the total matrix polysaccharide population.

More precise information about the polysaccharide composition and dynamics of the two mutant cell walls can be obtained by analyzing the peak height ratios between the low- and high-MW cell walls, taking into account sample amount differences. Because the two mutant cell walls have similar line widths, the ratios of the quantitative spectral intensities report the relative polysaccharide concentrations, whereas the ratios of the INEPT spectral intensities report the relative mobilities of the polysaccharides. Figure 2 shows that the two mutants have near-unity intensity ratios for most peaks in the quantitative spectra except for the 108 ppm Ara C1 peak, the 100 ppm GalA C1 peak, and the 54 ppm methyl-ester peak, whose intensity



**Figure 3.** 2D  $^{13}\text{C}$  J-INADEQUATE spectrum of the low-MW cell wall. The spectrum was measured at 296 K under 12 kHz MAS. Superscripts denote different subtypes of each monosaccharide. Boxed regions at double-quantum  $^{13}\text{C}$  chemical shifts of 160 and 170 ppm show the GalA C6 carbonyl signals and Rha-C6 signals, respectively.

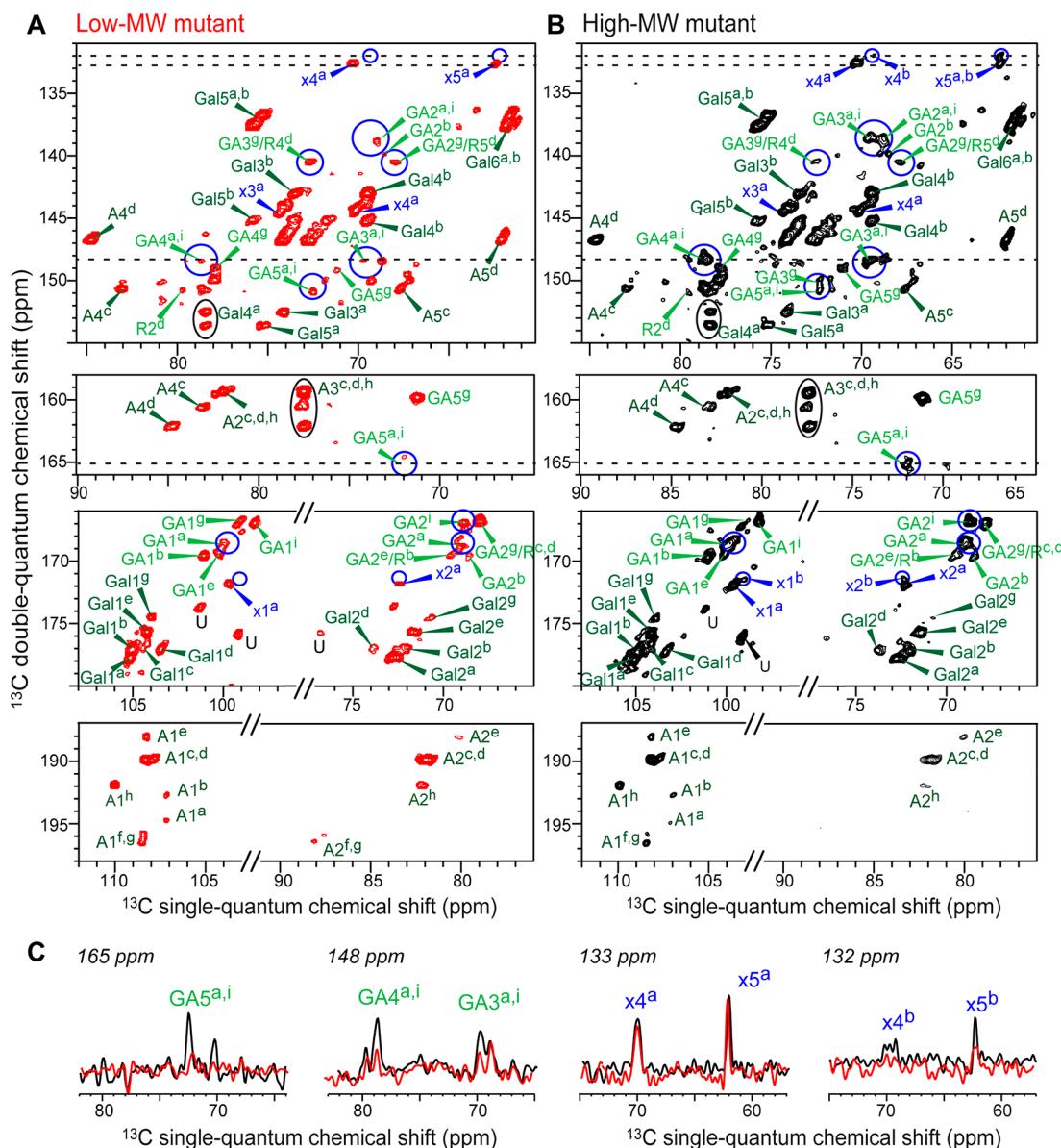
ratios are 1.2, 0.7, and 1.2. Because the 54 and 100 ppm peaks have varying contributions from neighboring  $^{13}\text{C}$  signals, we used spectral deconvolution to obtain more precise intensity values for these two peaks (Figure S1). After deconvolution, the relative intensities for the 54 ppm peak are 1.18, 1.00, and 1.14 for the low- and high-MW mutant and wild-type cell wall, respectively, whereas the relative intensities of the 100 ppm peak are 0.81, 1.00, and 1.05 for the three cell walls, respectively, in good agreement with the results without deconvolution.

Compared to the quantitative spectra, the INEPT intensity ratios between the low- and high-MW mutants deviate more substantially from 1, indicating different dynamics of the mobile polysaccharides that are selectively detected in the INEPT spectra. In particular, the Ara and Gal signals at 110 and 108 ppm have INEPT intensity ratios of 1.5 and 1.4, respectively (Figure 2), which are much larger than the corresponding quantitative intensity ratios, indicating that the low-MW mutant has much more mobile Ara side chains than the high-MW mutant. Compared to pectins, xyloglucan exhibits the opposite intensity trend: the 99.5 ppm Xyl C1 signal has an INEPT intensity ratio of less than 1.0, indicating that the low-MW  $PGXI^{AT}$  mutant has more rigid XyG compared to that of the high-MW cell wall.

The HG and RG-I backbones show an opposite mobility trend from the pectin side chains in the INEPT-detected spectra. The GalA and Rha signals such as the 69 ppm GalA C2 signal, the 18 ppm Rha C6 signal, and the 98.5 ppm GalA C1 signal, exhibit lower INEPT intensity ratios than the quantitative intensity ratios, indicating that the low-MW cell

wall has more rigid HG and RG-I backbones than that of the high-MW cell wall. Most prominently, the 100 ppm of HG GalA C1 peak (type a and e) has the lowest INEPT intensity ratio of 0.4 among all  $^{13}\text{C}$  signals (Figures 1B and 2), indicating that negatively charged HG is much more immobilized in the low-MW cell wall than in the high-MW mutant. The INEPT intensities of the 100, 54, and 18 ppm peaks indicate that the HG and RG-I backbones are the most rigid in the low-MW mutant, intermediate in the wild-type cell wall, and most dynamic in the high-MW mutant (Figure S2). Because the INEPT experiment detects only the most mobile population of matrix polysaccharides, these changes pertain to HG in the interfibrillar domain. The data suggest that the shorter HG chains in the low-MW cell wall are aggregated and/or constrained by other polysaccharides compared to the longer HG chains in the high-MW cell wall.

We also compared the two mutants with the wild-type cell walls using peak height ratios  $H_{\text{LowMW}}/H_{\text{Wild-type}}$  and  $H_{\text{HighMW}}/H_{\text{Wild-type}}$  (Figure S2). Again, we focus on the polysaccharide signals, whereas the  $^{13}\text{C}$  signals of lipids between 25 and 35 ppm, which do not originate from the cell walls, are not considered. Between the high-MW mutant and wild-type cell walls, and between the low-MW and wild-type cell walls, the intensity ratios of the quantitative spectra are close to 1 except for the 100 ppm nonmethylated HG signal. In contrast, the pectin side chain INEPT intensity ratios between the low-MW and wild-type cell walls are larger than those between the high-MW and wild-type cell walls. These results indicate that the three cell walls have similar polysaccharide compositions except for the negatively charged HG concentration, and the side



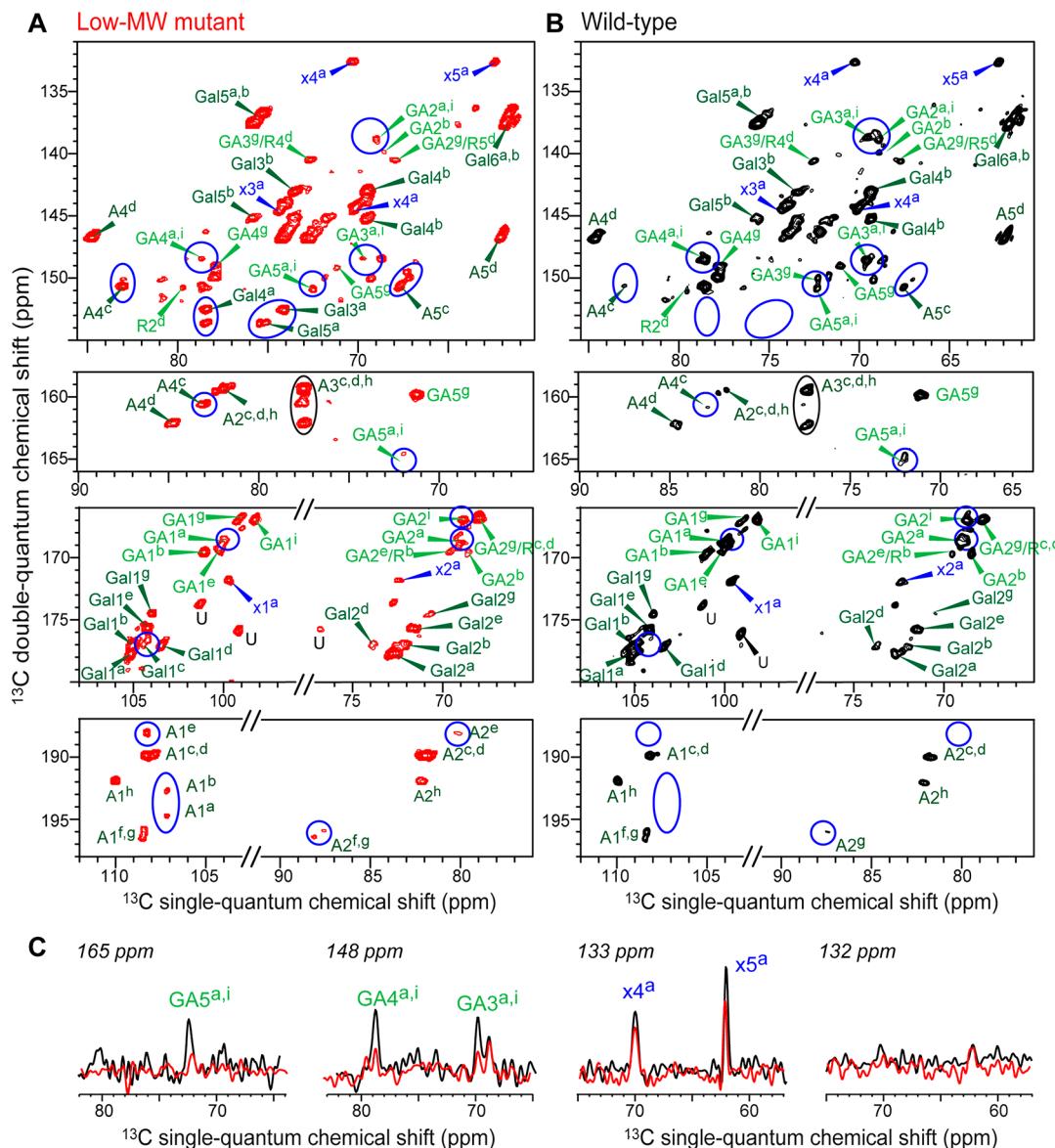
**Figure 4.** 2D  $^{13}\text{C}$   $J$ -INADEQUATE spectra of the low-MW (A) and high-MW (B) cell walls. The letter “U” denotes peaks whose assignments are unknown. On the basis of the C1–C2 chemical shifts, these peaks may be due to GalA, Rha, or Xyl. Blue circles highlight regions of significant intensity differences. (C) Selected 1D  $^{13}\text{C}$  cross sections show intensity differences of matrix polysaccharide signals between the two cell walls. The intensities of Xyl-a and Xyl-b are reduced by  $\sim 10\%$  and  $\sim 70\%$  respectively, and the intensity of GalA-i is lower by 10–70% in the low-MW cell wall (red). The cross sections are plotted after multiplying the low-MW spectrum by 0.64 to account for sample amount differences.

chains are similarly mobile between high-MW and wild-type cell walls but much more mobile in the low-MW cell wall.

**Compositions of HG and XyG from 2D  $^{13}\text{C}$  INADEQUATE Spectra.** To fully resolve the matrix polysaccharide signals, we measured 2D  $^{13}\text{C}$ – $^{13}\text{C}$   $J$ -INADEQUATE spectra (Figure 3), which use  $^{13}\text{C}$ – $^{13}\text{C}$   $J$ -coupling to transfer  $^{13}\text{C}$  coherence and correlate the sum of the chemical shifts of two directly bonded  $^{13}\text{C}$  spins with the chemical shift of each  $^{13}\text{C}$ .<sup>22,52</sup> In addition to scalar-coupling based polarization transfer, the 2D spectra were measured using direct  $^{13}\text{C}$  polarization and a short recycle delay of 2 s to preferentially detect the signals of mobile polysaccharides. Approximately 90% of the matrix polysaccharides were observed in the 2D INADEQUATE spectra, as estimated based on the fact that 1D  $^{13}\text{C}$  DP spectra measured with a 2 s recycle delay showed  $\sim 90\%$

of the matrix polysaccharide signals compared to the quantitative spectra.

Figure 3 shows the 2D INADEQUATE spectrum of the low-MW cell wall. Similar to previously studied cell walls, the spectrum is dominated by pectin signals with narrow line widths of 0.4–0.8 ppm followed by a small number of xylose peaks. Multiple linkages were detected for the matrix polysaccharides, including eight types of Ara, six types of Gal, five types of GalA, four types of Rha, and two types of Xyl (Table 1). Comparison of specific spectral regions between the low- and high-MW samples (Figure 4) indicates that the two mutants differ mainly in peak intensities rather than chemical shifts. For example, type-a, i GalA intensities are lower by 10–70%, and type-a and type-b Xyl intensities are lower by  $\sim 10\%$  and  $\sim 70\%$  in the low-MW cell wall (Figure 4C). Because the Rha content is the same between the two mutants,<sup>43</sup> these GalA

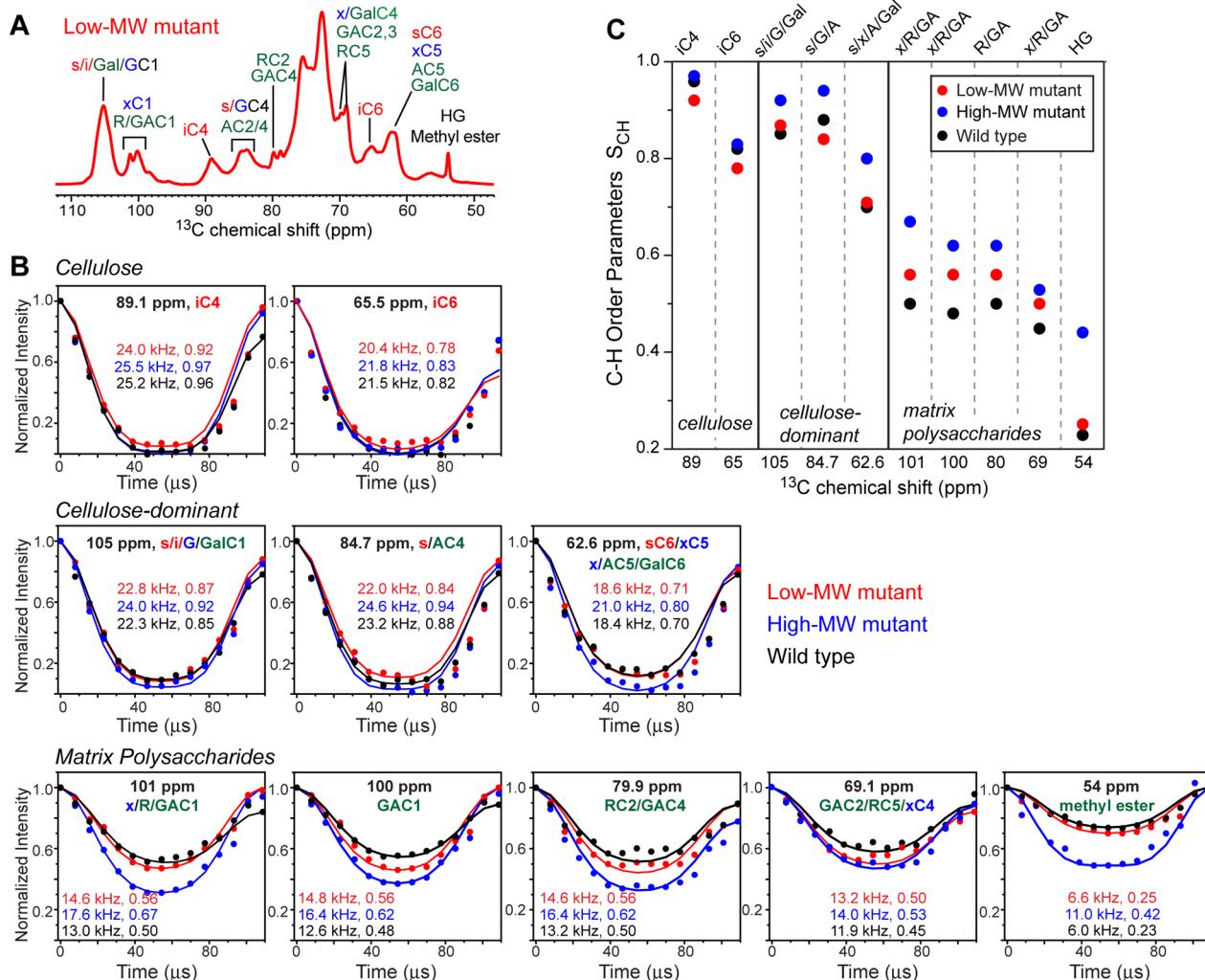


**Figure 5.** 2D  $^{13}\text{C}$   $J$ -INADEQUATE spectra of the low-MW (A) and wild-type (B) cell walls. Blue circles highlight regions of significant intensity differences. The letter “U” denotes peaks whose assignments are unknown. The low-MW sample shows weaker GalA-i and Xyl-b signals and weaker or missing Ara-a,b,c,e,f and Gal-a side chain signals. (C) Selected 1D  $^{13}\text{C}$  cross sections showing intensity differences for matrix polysaccharide signals between the two cell walls. The intensity of GalA-i is lower by 30–60% in the low-MW cell wall (red), and the intensity of Xyl-a is reduced by ~30%. The cross sections are plotted after multiplying the low-MW spectrum by 0.67 to account for sample amount differences.

intensity changes can be attributed to HG, indicating that the low-MW  $\text{PGX1}^{\text{AT}}$  cell wall contains less dynamic HG and XyG than the  $\text{pgx1-2}$  cell wall, consistent with the INEPT data. The GalA signals in the INADEQUATE spectra are similar between wild-type and high-MW cell walls, (Figures 4, 5), also consistent with the quantitative spectra (Figure 1A). Finally, both mutants show higher Ara and Gal intensities than those of the wild-type cell wall, consistent with the quantitative  $^{13}\text{C}$  spectra (Figure 1A), indicating that the mutants have more RG-I side chains than the wild-type cell wall.

**C–H Dipolar Order Parameters.** To investigate whether  $\text{PGX1}$  mutation affects the dynamics of more rigid polysaccharides that are not well detected by the INEPT and INADEQUATE experiments, we measured  $^{13}\text{C}$ – $^1\text{H}$  dipolar couplings using the dipolar chemical-shift (DIPSHIFT) correlation experiment (Figure 6).<sup>54,55</sup>  $^1\text{H}$ – $^{13}\text{C}$  cross-polarization (CP) was used to create the initial  $^{13}\text{C}$  magnetization;

thus, the experiment selectively detects the rigid wall polysaccharides, including cellulose and ~40% of the matrix polysaccharides, as estimated from comparisons of the CP and quantitative spectra. Figure 6B shows the C–H dipolar dephasing curves and the corresponding order parameters of all major polysaccharides. As expected, cellulose exhibits near rigid-limit order parameters for all three cell walls. For matrix polysaccharides, the low-MW  $\text{PGX1}^{\text{AT}}$  cell wall shows moderately larger order parameters than wild-type cell walls, whereas the high-MW  $\text{pgx1-2}$  mutant has much larger order parameters than both control and low-MW cell walls (Figure 6C). The largest difference between the two mutants is found for the 101, 84.7, and 62.6 ppm peaks as well as the 54 ppm methyl ester peak. Because the CP-DIPSHIFT spectrum preferentially detects more rigid pectins, these results indicate that the pectin–cellulose interaction is weaker in the control and low-MW  $\text{PGX1}^{\text{AT}}$  cell walls than in the high-MW mutant.



**Figure 6.**  $^{13}\text{C}$ – $^1\text{H}$  dipolar-doubled CP-DIPSHIFT data of the low-MW (red), high-MW (blue), and wild-type *Arabidopsis* cell walls (black). (A)  $^{13}\text{C}$  CP spectrum of the low-MW mutant showing various  $^{13}\text{C}$  signals of interest. (B)  $^{13}\text{C}$ – $^1\text{H}$  dipolar dephasing curves of cellulose signals (top row), cellulose-dominant signals (middle row), and matrix polysaccharides signals (bottom row). The data were measured at 296 K under 9.2 kHz MAS. Each panel in (B) gives the best-fit dipolar couplings (kHz) after taking into account the FSLG scaling factor of 0.577. The order parameters  $S_{\text{CH}}$  are also given. (C) C–H dipolar order parameters. The high-MW cell wall is more immobilized than the low-MW cell wall. The difference is the largest for the 101 ppm R/GA C1 signal, the 62.6 ppm s/x/A/Gal signal, and the 54 ppm methyl ester signal.

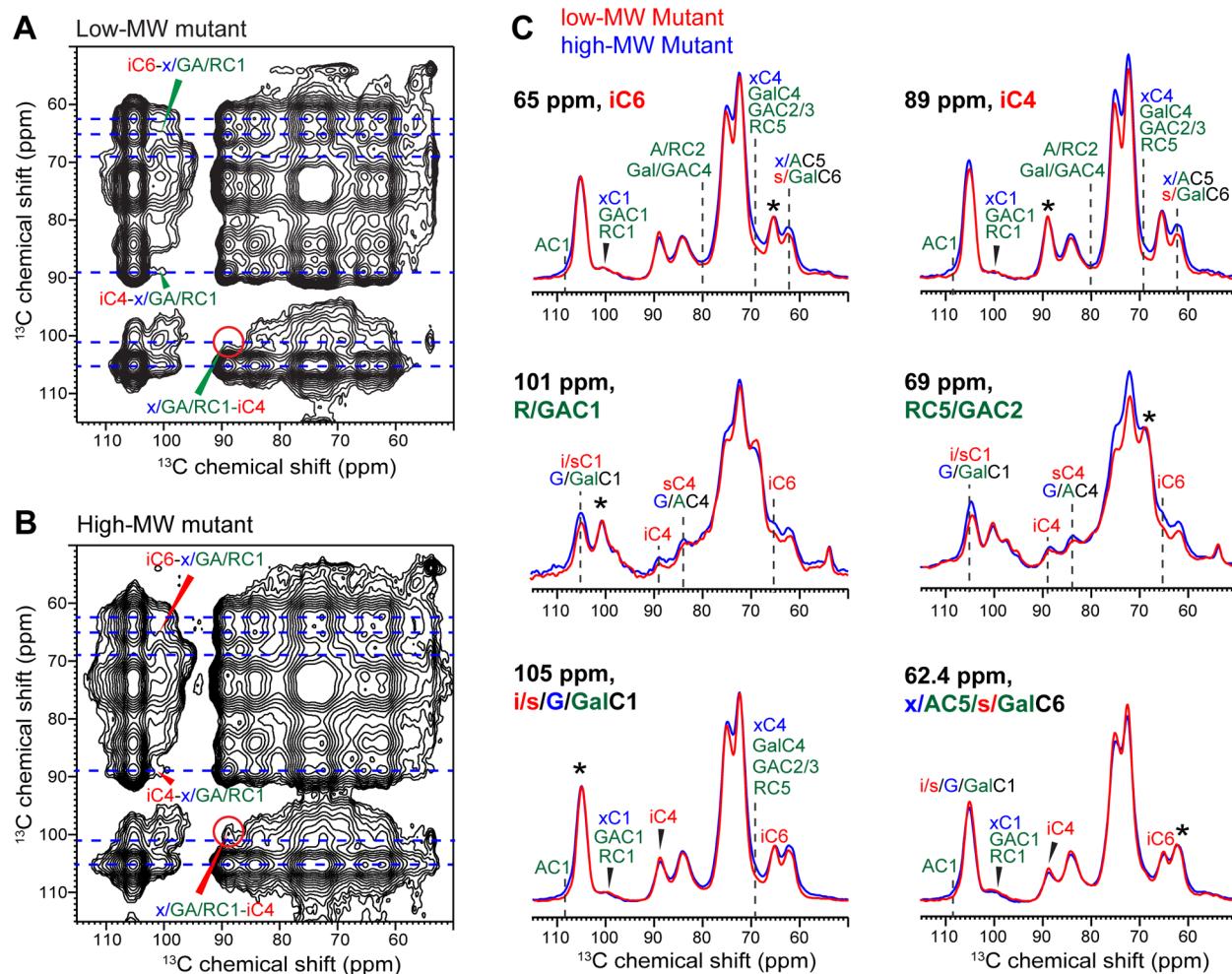
In particular, the high-MW mutant exhibits a large order parameter of 0.42 for the 54 ppm methyl ester peak, suggesting that the three-site jump motions of  $\text{CH}_3$  groups are significantly impeded by polysaccharide interactions.

**Pectin–Cellulose Proximities from 2D  $^{13}\text{C}$ – $^{13}\text{C}$  Correlation Spectra.** To further investigate whether pectin–cellulose interactions differ between the two mutants, we measured 2D  $^{13}\text{C}$  PDSD spectra using a long  $^{13}\text{C}$  mixing time of 1.5 s (Figure 7). The spectra exhibit extensive pectin–cellulose cross peaks such as the interior cellulose C4 and C6 correlations to the C1 peak of Xyl, GalA, or Rha at 89 and 101 ppm and at 65 and 101 ppm, respectively. Importantly, in the 89 and 65 ppm cross sections of interior cellulose, the low-MW cell wall shows weaker pectin cross peaks than those of the high-MW mutant. Examples include the 108 ppm peak of Ara C1, the 80 ppm peak of Ara/Rha C2 and Gal/GA C4, and the 69.7 ppm peak of a mixture of pectin and Xyl carbons (Figure 7C). Conversely, in the 101 and 69 ppm cross sections of pectin carbons, the low-MW cell wall exhibits weaker cellulose cross peaks at 89 and 65 ppm than those of the high-MW

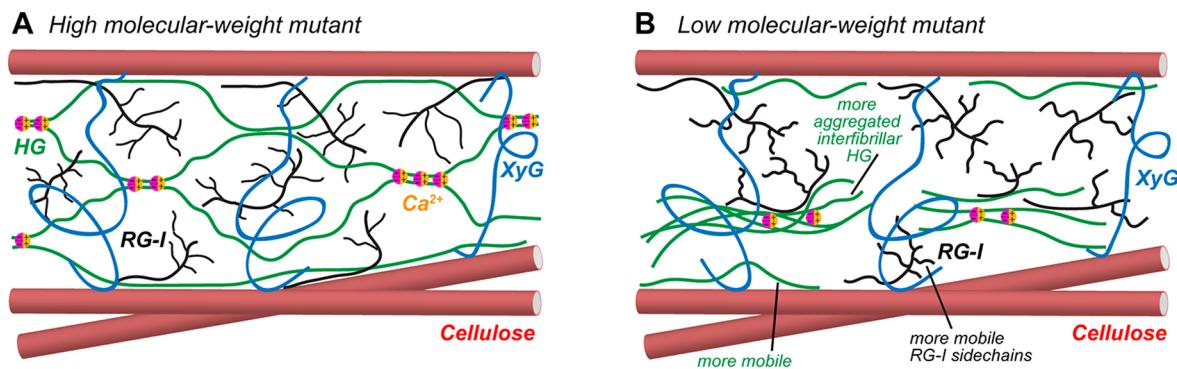
mutant. These results indicate that the low-MW  $PGX1^{AT}$  cell wall has weaker interactions between cellulose and the rigid domain of HG and RG-I, which is consistent with the higher HG and RG-I backbone mobilities of the  $PGX1^{AT}$  cell wall seen in the CP-DIPSHIFT data.

## DISCUSSION

These SSNMR data, together with previous biochemical characterization of  $PGX1$  mutants,<sup>43</sup> shed light on how pectin–cellulose interactions and pectin conformational dynamics correlate with the extensibility of primary cell walls.<sup>28,60</sup>  $PGX1$  mutations alter HG molecular weight and either promote ( $PGX1^{AT}$ ) or limit ( $pgx1-2$ ) cell expansion, hypocotyl elongation, and adult plant growth.<sup>43</sup> These changes in cell and organ growth may be attributed to changes in the characteristics and behaviors of HG: in  $PGX1^{AT}$  cell walls, the increased polygalacturonase activity is expected to degrade regions of HG chains with low esterification, which should shorten HG chains, attenuate the calcium-cross-linked pectin network, and in turn cause a looser wall. The reverse would be



**Figure 7.** 2D <sup>13</sup>C PDSD spectra of the (A) low- and (B) high-MW mutant to detect intermolecular cross peaks. The spectra were measured at 253 K under 10 kHz MAS using a 1.5 s spin diffusion mixing time. (C) Selected 1D <sup>13</sup>C cross sections normalized by the diagonal peaks (asterisks). Dashed lines indicate pectin–cellulose cross peaks of interest.



**Figure 8.** Models of the different polysaccharide dynamics and interactions in the (A) high- and (B) low-MW mutant cell wall. In both cell walls, HG (green) and RG-I (black) have a rigid cellulose-proximal domain and a mobile interfibrillar domain. The low-MW mutant shows more dynamic pectins near the cellulose microfibril (red) but more rigid interfibrillar HG and RG backbones, suggesting increased aggregation due to reduced calcium cross-linking. The RG-I side chains are more mobile in the low-MW cell wall. These differences correlate with enhanced growth of the low-MW mutant.

expected for the *pgx1-2* mutant, where a reduction in total polygalacturonase activity is expected to give rise to longer HG chains with low esterification, which may then cross-link via calcium, stiffen the wall, and diminish wall expansibility and cell growth. Pectinase treatment can restore expansin activity in

cucumber hypocotyls,<sup>61</sup> implying that pectins also limit wall expansibility in that species.

The solid-state NMR data shown here support some aspects of the hypothesis presented above but also reveal unexpected structural and dynamical features of the pectin network. Figure

8 summarizes the findings from the SSNMR data and compares the polysaccharide mobilities and intermolecular interactions between the two mutant cell walls. The concentrations of most polysaccharides are similar between the two mutant walls with the exception that the low-MW *PGX1<sup>AT</sup>* cell wall has significantly lower amounts of negatively charged carboxyl groups and higher esterification of HG. The enrichment of methyl-esterified HG in *PGX1<sup>AT</sup>* walls may result from the resistance of esterified HG to degradation by PGX1 and other polygalacturonases. Alternatively, it may arise from higher levels of synthesis and/or exocytosis of new highly esterified HG due to pectin degradation and oligogalacturonide release, although the similar levels of total pectic HG content between *PGX1<sup>AT</sup>* and wild-type walls (Figure 1A) argue against this idea. A third possibility is that the increased polygalacturonase activity and resulting higher levels of oligogalacturonides<sup>62</sup> may upregulate HG esterification during HG synthesis or downregulate HG de-esterification by pectin methyl-esterases.

The different HG esterifications are coupled with complex mobility changes between the low- and high-MW mutant cell walls. The mobilities change in opposite directions for the dynamic and rigid domains. For the dynamic pectin domain between cellulose microfibrils, the low-MW cell wall has more immobilized HG and RG-I backbones but more dynamic arabinan and galactan side chains compared to those of the high-MW cell wall. We attribute the backbone immobilization to partial HG aggregation in the interfibrillar space. This could occur via one of two mechanisms. First, the increased HG esterification in the low-MW mutant would be expected to reduce calcium cross-linking, thus hindering the formation of an open, well-spaced pectin network in the interfibrillar space (Figure 8B). Alternatively, the lower negative charge of HG might increase intermolecular interactions between HG chains or between HG and RG-I or RG-II,<sup>10,12</sup> thus facilitating aggregation. HG aggregation can in turn create open regions in which RG-I side chains can undergo larger-amplitude motions. In contrast to the mobile pectin domain, the CP-DIPSHIFT spectra show that the rigid pectin domain that interacts with cellulose becomes less rigid in the low-MW cell wall and shows weaker cross peaks with cellulose in 2D <sup>13</sup>C-<sup>13</sup>C correlation spectra. This could result from less cross-linking of HG in the vicinity of cellulose or shorter HG chains that have limited interactions with cellulose. Taken together, these data suggest that the enhanced cellular growth observed in the low-MW *PGX1<sup>AT</sup>* mutant may result from a less-cross-linked pectin network and weaker pectin-cellulose interactions, both of which can facilitate wall loosening and expansion.

These SSNMR data imply that it is the pectin backbones rather than side chains that interact with cellulose to regulate wall extensibility, even though *in vitro* binding assays found that the neutral arabinan and galactan side chains are better than the charged pectin backbones in binding cellulose.<sup>16,36,63</sup> That RG-I side chains do not play a significant role in cellulose interactions in *planta* is supported by the fact that Ara and Gal show strong signals in the INEPT spectra but negligible intensities in the CP spectra. The lack of extensive interactions between pectin side chains and cellulose may be due to conformational disorder of the side chains, whereas the stronger interactions of the HG backbone with cellulose may be mediated by favorable HG backbone conformations. *PGX1* overexpression, by reducing the molecular weight and increasing the esterification of HG, simultaneously weakens HG-cellulose interactions and causes partial aggregation of the interfibrillar HG. It remains to be

determined how HG synthesis, esterification, and de-esterification in the wall might be altered in *PGX1* mutants to compensate for changes in HG autodegradation. We hypothesize that this could occur via wall-integrity-sensing pathways that detect HG degradation products such as oligogalacturonides.<sup>64</sup> Pectin degradation might indirectly affect wall composition and architecture by activating defense responses and/or wall integrity-sensing pathways.<sup>62</sup> This is an especially attractive idea if *PGX1* acts as an endopolygalacturonase, which seems likely from the ~2.3-fold changes in HG molecular weight observed in the *PGX1<sup>AT</sup>* cell walls,<sup>43</sup> and produces high levels of oligogalacturonides. When expressed heterologously, *PGX1* possesses at least 10-fold higher polygalacturonase activity than three other *Arabidopsis* polygalacturonases,<sup>43,65</sup> and total polygalacturonase activity increased by ~50% in *PGX1<sup>AT</sup>* leaves and decreased by ~50% in *pgx1-2* leaves relative to wild-type.<sup>43</sup> It is thus somewhat surprising that only 2–3 additional cuts per HG molecule appear to occur. Experiments to detect oligogalacturonide levels and wall-integrity activation in *PGX1* mutants should be informative for answering these questions.

The fact that a decrease and increase of HG molecular weight in the *PGX1<sup>AT</sup>* and *pgx1-2* cell walls causes enhanced and diminished hypocotyl elongation, respectively, suggests that, in the wild-type cell wall, homogalacturonan–cellulose interactions and pectin network formation play an important role in constraining cell growth. We speculate that this constraining effect may be directional to control the anisotropy of cell expansion.<sup>66</sup> Finally, the INEPT spectra show that XyGs become more rigid in the low-MW mutant, suggesting that XyG may compensate for weaker pectin–cellulose interactions by binding more extensively to cellulose as a larger microfibril surface area becomes available. Future biomechanical, genetic, and structural analyses should help to delineate how pectin–cellulose and XyG–cellulose interactions might work in a concerted fashion to regulate wall dynamics during plant growth.

## CONCLUSIONS

The current solid-state NMR data on the pectin mutant cell walls reveal the importance of homogalacturonan–cellulose interactions and pectin network formation in regulating cell wall growth. The low-MW mutant, with higher *PGX1* activity, lower HG molecular weight, and longer hypocotyls, exhibits distinct polysaccharide compositions, dynamics, and intermolecular proximity to cellulose from control cell walls. The low-MW mutant has lower galacturonic acid, xylose, and carboxylate contents and higher methyl esterification levels. The increased esterification leads to aggregation of HG and RG-I backbones in the interfibrillar space, likely because lower negative charges or reduced steric constraints hinder the formation of a well-spaced pectin network. The increased pectin backbone aggregation is correlated with increased mobility of the RG side chains, likely due to increased space for side chain motion. Further, the low-MW cell wall shows higher mobility of pectins that interact with cellulose. These results show that growth enhancement of the low-MW mutant is correlated with increased esterification, reduced cross-linking, and increased aggregation of interfibrillar pectins and weaker HG–cellulose microfibril interactions.

## ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge on the [ACS Publications website](#) at DOI: [10.1021/acs.biromac.7b00888](https://doi.org/10.1021/acs.biromac.7b00888).

Spectral deconvolution of selected signals and comparison of  $^{13}\text{C}$  spectral intensities between the mutants and wild-type cell walls ([PDF](#))

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### Notes

The authors declare no competing financial interest.

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## ABBREVIATIONS

Ara, A, arabinose

CW, cell wall

CP, cross-polarization

DP, direct polarization

Gal, Galactose

GalA, GA, galacturonic acid

HG, homogalacturonan

INEPT, insensitive nuclei enhanced by polarization transfer

MAS, magic-angle spinning

PDSD, proton-driven  $^{13}\text{C}$ – $^{13}\text{C}$  spin diffusion

SSNMR, solid-state nuclear magnetic resonance

MW, molecular weight

PGX1, POLYGALACTURONASE INVOLVED IN EXPANSION1

RG-I, rhamnogalacturonan I

Rha, R; rhamnose

XyG, xyloglucan

Xyl, x, xylose

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